

Ruslan I. Dmitriev, PhD, Associate Professor
06-November-2026

Vision for the Society:

I am excited to apply for the Council member position at Histochemical Society. I hope that I can help this society to evolve by supporting its scientific mission and excellence (my inter-disciplinary research expertise in dynamic visualisation of biochemical and structural organisation of cells, tissues and organs using FLIM and PLIM bio-, chemical and nanosensors), harmonisation and standardisation of imaging approaches across the globe (expertise in teaching and mentoring in imaging methods, e.g. inter-university courses in Flanders and EU, including FEBS imaging course) and connecting research expertise between the US-based researchers and overseas, via participation in Flanders and Euro-BioImaging networking activities. Being a research group leader (Faculty of Medicine & Health Sciences, Ghent University, Belgium), with the primary focus on fluorescence lifetime imaging microscopy-assisted analysis and manipulation of organoids, 3D tissue constructs and drug delivery models, I am also involved in Ghent Light Microscopy Core, Flanders BioImaging consortium (connecting the centres of excellence in KU Leuven, UHasselt, Ghent, Antwerp and VIB with Euro- and Global BioImaging consortia), EU-UK wide Horizon consortium 'FLIMagin3D', and numerous international collaborations outside of the EU (e.g. HHMI Janelia research campus, University of Nottingham, Manchester and King's College London).
I am convinced that my multi-national experience (born in Soviet Union, grown up in Russian Federation, matured as a scientist in Ireland, performed research in more than 3 countries across the globe) is a strong 'plus' in understanding and addressing minority, equity and diversity issues as well as internationalisation of research. Seeing HCS as a premier society in histochemistry and cell-based imaging, I hope that my research and educational experience, vision and growing research network can help supporting its international & global missions.

Please find my recent bio-sketch in the following pages.

BIOGRAPHICAL SKETCH

NAME: Ruslan I. Dmitriev, PhD

POSITION TITLE: Associate Professor, Head of Tissue engineering and Biomaterials Imaging Group; Chairman of Ghent Light Microscopy Core (GLIM)

EDUCATION/TRAINING

INSTITUTION AND LOCATION	DEGREE	Start Date	Completion Date	FIELD OF STUDY
Moscow State Academy of Fine Chemical Technology, Moscow, Russia	B.Sc. (distinction)	09/1998	06/2002	Chemistry and Technology
Moscow State Academy of Fine Chemical Technology, Moscow, Russia	M.Sc. (distinction)	09/2002	06/2004	Chemistry and Technology of Biologically Active Compounds
Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry, Russian Academy of Sciences, Moscow, Russia	Ph.D.	09/2004	05/2008	Bioorganic chemistry
DCU Ryan Academy, Dublin, Ireland	Certificate of completion	01/2013	06/2013	'Get started technology venture' Course for entrepreneurs

A. Personal Statement

I am a microscopist focusing on developing bioconjugate, nanoparticle- and scaffold-based biosensors for live cell 4D imaging. My primary research interest is in advancing **quantitative, live and multi-parameter** fluorescence (FLIM) and phosphorescence (PLIM) lifetime imaging microscopies. These approaches are instrumental for metabolic, O_2, FRET and many other dynamic imaging modalities, applicable to 3D organoids & related tissue models. During my doctoral and postdoctoral training I gained interdisciplinary experience in studies of cell metabolism, hypoxia and development of phosphorescent high-performance biosensors. In 2014-2019 I formed Metabolic Imaging Group at University College Cork, followed up by the industry fellowship at Agilent technologies (2019-2020), before joining Ghent University (2020) for tenure-track Professor position. My current ambitions are in promoting live 3D-FLIM approaches to **visualize the single cell metabolism & oxygenation within organoids and *in vivo* models**. To date, I've gained research leadership in the areas of quenched phosphorescence-based detection of molecular oxygen and multi-parameter FLIM of **spheroids, neural and intestinal organoids**, which is evident by number of reviews, editorship of 2 books (2017, 2018), organizing the research meetings and the society membership (HCS, ASCB, BPS). My industrial experience is in generating 3 invention disclosures, 1 patent application and collaborations with SME and MNE such as Luxcel Biosciences (now part of Agilent Technologies), Lambert Instruments, Becker & Hickl GmbH, Leica Microsystems and Rousselot. Since 2026, I am also serving as a chairman at Ghent Light Microscopy Core, part of Flanders Bio-Imaging and Euro-BioImaging consortia. Please find below a list of my **on-going** research projects:

1. **Inter-university special research fund (VLIR-iBOF)** grant 'MINDFUL' IBOF/25/28. *Lipid metabolism directed crosstalk between microglia and neurons as driver of neurodegeneration in MS and AD.* Promotor at UGent. **2025-2028**
2. **Special Research Fund (BOF) Interdisciplinary research project** (BOF/IOP/2022/058) (Co-PI with Prof. S. Vergult): *IMAGIBRAINDD: IMAGIng-assisted human BRAIn organoid engineering for studying and treating NeuroDevelopmental Disorders*. **2022-2028**
3. **HORIZON-MSCA-2021-DN-01** (101073507) fliMAGIN3D (Co-PI, 12 partners): *Doctoral Network for a Shared Excellence of Fluorescent Lifetime Imaging Microscopy in Biomedical Applications*. 3 PhD students for my team. **2023-2026**
4. **Research Foundation Flanders (FWO) Medium-size infrastructure grant** (I004124N) Two-photon upgrade of Stellaris 8 FLIM microscope, with 22 co-promotors, 660,426.76 euro. **2024-2028**

Please find below a list of ***four of my significant publications*** directly related to this research program:

1. Mamillapalli VNK, Okkelman IA, Akhssas F, Sambucari G, Debruyne AC, Beniwal D, Bucher C, Donato MD, Dmitriev RI Filatov MA: Hydration-Gated Photoinduced Electron Transfer in BODIPY Dyes. ***Angewandte Chemie International Edition*** 2026, **n/a**(n/a):e3726851.
2. Okkelman IA, Zhou H, Borisov SM, Debruyne AC, Lefebvre AEYT, Leomil Zoccoler M, Chen L, Devriendt B, Dmitriev RI: Visualizing the internalization and biological impact of nanoplastics in live intestinal organoids by Fluorescence Lifetime Imaging Microscopy (FLIM). ***Light: Science & Applications*** 2025, **14**(1):272.
3. Vanhove L, Van Gansbeke T, Devriendt B, Van der Meeren R, Dmitriev RI, Okkelman IA: Lack of biochemical signalling in GelMA leads to polarity reversion in intestinal organoids independent from mechanoreciprocity**.** ***Journal of Tissue Engineering*** 2025, **16**:20417314251345000.
4. Debruyne AC, Okkelman IA, Heymans N, Pinheiro C, Hendrix A, Nobis M, Borisov SM, Dmitriev RI: Live Microscopy of Multicellular Spheroids with the Multimodal Near-Infrared Nanoparticles Reveals Differences in Oxygenation Gradients. ***ACS Nano*** 2024, **18**(19):12168-12186.

B. Positions, scientific appointments, and Honors

Employment (positions held):

2025-present – Associate Professor, Department of Human structure and repair, Ghent University, Ghent, Belgium
2020-2025 – Assistant Professor (tenure track), Department of Human structure and repair, Ghent University, Ghent, Belgium
2014-2020 (full-time) – Research Fellow/ Starting Investigator, School of Biochemistry, University College Cork, Ireland
2018-2020 (part-time) – Chief Researcher, Institute for Regenerative Medicine, Sechenov Medical University, Moscow, Russia
2014-2014 – Senior Postdoctoral researcher (Tyndall National Institute – University College Cork, IPIC project)
2008-2012 – Postdoctoral Researcher, School of Biochemistry, University College Cork
2004-2008 – Junior Scientist (part-time), Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry, Russian Academy of Sciences

Other appointments:

2022-present – Member of the steering committee 'Ghent Light Microscopy Core' (GLIM), **chairman** (since 2026)

2020-present – Member of the 'Ghent Gut Inflammation Group' (GGIG) https://www.ggig.be
2019-2020 – Industry fellow at the Agilent Technologies (Agilent, Little Island, Cork)
2004-2008– Analyst (part time), Newcastle Technical Centre of Procter & Gamble, Moscow, Russia (Affiliate lab)

Honors, professional membership and books editorship:

- **Early Career Investigator** Award (Histochemical Society)
- 2012-2025: Continuously secured **16** research grants, total worth ~9.8M euro (see section D)
- 2005-2024: Received **15** travel grants to attend the international conferences and support research visits, from the American Society of Biochemistry and Molecular Biology (ASBMB, graduate student and postdoctoral travel awards for Experimental Biology symposia; USA; 2007, 2011, 2012, 2014, 2015), American Society for Cell Biology (ASCB, postdoctoral travel award for Annual Meeting; USA; 2011, 2017, 2019, 2022), Society of General Physiologists (USA; 2005), Biophysical Society (2020), Research Foundation Flanders (FWO, 2022, 2023), HHMI Janelia Visitor Scientist Program (2024), Biophysical Society Bridge Funding Award (2026) and Histochemical Society Travel Award (2026).
- Edited, co-edited and authored **3 books:**

1. Papkovsky DB, Dmitriev RI (eds.): Quenched-phosphorescence Detection of Molecular Oxygen: Applications in Life Sciences: The Royal Society of Chemistry; 2018. ISBN 978-1-78801-175-4.
2. Dmitriev RI (ed.): Multi-Parametric Live Cell Microscopy of 3D Tissue Models: Springer International Publishing AG 2017; 2017. ISBN 978-3-319-67357-8.
3. Papkovsky DB, Zhdanov AV, Fercher A, Dmitriev RI, Hynes J: Phosphorescent Oxygen-sensitive probes. Springer Briefs, Basel, 2012. ISBN 978-3-0348-0524-7.

- Member of Histochemical Society (since 2022), American Society for Cell Biology (since 2011, in 2017-18: ambassador), Tissue engineering and regenerative medicine (TERMIS, Invited speaker, co-organiser of symposia at the meetings, since 2016).
- (2022-onwards) Co-editor of the methods collection at Journal of Visualized Experiments (**JoVE**): Quantitative live cell imaging of 3D models: https://www.jove.com/methods-collections/902
- 2019 – 'Best paper award' (Irish Photonic Integration Centre, Cork, Ireland)

Teaching responsibilities

2020-present – teaching (lecturer-in charge and co-lecturer) M.Sc. courses in biomedical science and engineering (Ghent University), Tissue engineering, Biomaterials and Tissue engineering, Image to numbers (6 in total, 29 credits). Major in Tissue Engineering M.Sc. in Biomedical Sciences Program responsible
2020-present – invited lecturer at Trinity College Dublin for M.Sc. Advanced imaging course

Peer-review and grant review activities

Referee for research journals https://www.webofscience.com/wos/author/record/1752038
2017 – ACS (American Chemical Society) Reviewer Lab certificate
2015 – Certificate of 'Outstanding contribution in reviewing'. Sensors and Actuators B: Chemical journal (Elsevier)
Evaluation of grant applications (Swiss National Fund SNF, 2018, Dutch Research Council N.W.O., 2019, IBiSA France, 2022). Reviewer of nominations Canada Research Chairs (2022), Israel Science Foundation (ISF, 2025), Frans van Cauwelaertprijs KVAB (2025).

Session chair / Program committee

2026 - Program committee “Translational Fluorescence Lifetime–Based Molecular Imaging” (BO111, **Photonics West,** SPIE conference)

2025 – **Co-organizer** of the Macromolecular Chemistry & Soft Matter Connects international symposium (Ghent, 18-19 December 2025, with Peter Dubruel and Bruno De Geest)

2025 **ESMI EMIM 2026 conference** reviewer / programming chair ‘Theranostics, Multimodal Agents | Probes/Reporter Genes”

2023 **TERMIS-EU Manchester** (Mar-2023) / Imaging symposium / Co-organizer, co-chair and invited speaker

2022 **TERMIS-EU Krakow /** Imaging symposium / Co-organizer and co-chair

2021 **TERMIS-World Congress** / Online-Maastricht NL / Section ‘Advanced spatiotemporal imaging for regenerative medicine’ (Co-organizer and co-chair).

2020-cancelled due to COVID-19 pandemic **TERMIS-EU 2020 Meeting** / Manchester, UK / Oral presentation (invited speaker, **section organizer and co-chair**)

2019 **Sechenov International Biomedical Summit** / Moscow, Russia / **Session organizer and Chair**

2018 **American Society for Cell Biology (ASCB)** annual meeting / poster abstract programming

2018 **TERMIS – World Congress** / Kyoto, Japan / **Organizer and Co-Chair of** ‘Multi-parametric quantitative imaging of live cells in 3D tissue models’ symposium

2016 **TERMIS-EU meeting** / Uppsala, Sweden / **Co-Chair** of ‘Non-Destructive Imaging in TERM’ session

Invited talks

2027 Photonics West (San Francisco, 1-2 Feb 2027) (forthcoming)

2026 Inter-University Course on Advanced Light Microscopy (iCALM), October 2026

2026 Engineering life: advances in bioengineering and drug delivery. Joint TU/e-UGhent symposium (22 October 2026, Ghent)

2026 FEBS Practical Course ‘Functional Imaging of Cellular Dynamics’ / Amsterdam, NL (instructor)

2026 BELSACT Meeting 2026 (Ghent, Mar-2026)

2026 HiLight symposium (Brunel University, Uxbridge, UK, 4-5 Mar 2026)

2025 FLIMposium (King’s College London, launch of new Wellcome Trust Bio-Imaging centre, May-2025)

2024 SPAOM meeting (Toledo, Nov-2024)

2024 4th Inflammation & Hypoxia symposium (Munster, Sep-2024)

2024 FEBS Practical Course ‘Functional Imaging of Cellular Dynamics’ / Amsterdam, NL

2024 Belgian society for animal cell technology 2024 scientific meeting / Gosselies, Belgium

2024 FLIMagin3D consortium 1st networking event / Dublin, Ireland

2023 Albany Medical College / Albany, NY (seminar)

2023 VIB Cancer Biology Tech talk 2022-23 / Leuven, Belgium (seminar)

2022 Leuven Advanced Microscopy Symposium 2022

2022 32nd Annual Meeting of the German Society for Cytometry (DGfZ) / Berlin

2019 Sechenov International Biomedical Summit / Moscow, Russia

2018 Sechenov International Biomedical Summit / Moscow, Russia

2017 FLIM workshop / Prague, Vestec, Czech Republic

2016 TERMIS-EU meeting / Uppsala, Sweden

2006 1st International Young Scientists’ Conference / Kharkiv, Ukraine 2nd prize for best presentation

C. Contributions to Science

Addressing dynamics and heterogeneity of hypoxia and metabolic gradients in multicellular spheroid and organoid models.

My team focuses on multiplexed O_2 imaging of cancer cell evolution in 3D, developing 'metabolic FLIM' pipeline for addressing neurodevelopmental disorders using neural organoid models, and mechanical & biochemical interactions of polymer (bio)materials with small intestinal organoids. My team pioneered monitoring oxygenation in neurospheres (2013), small intestinal organoids (2017) and were among the first to visualize metabolic heterogeneity (affected by the medium & nutrient composition) of tumor spheroids & small intestinal organoids (2020-2024).

1. Dmitriev RI, Zhdanov AV, Nolan YM, Papkovsky DB: Imaging of neurosphere oxygenation with phosphorescent probes. ***Biomaterials*** 2013, **34**(37):9307-9317.
2. Okkelman IA, Foley T, Papkovsky DB, Dmitriev RI: Live cell imaging of mouse intestinal organoids reveals heterogeneity in their oxygenation. ***Biomaterials*** 2017, **146**:86-96.
3. Okkelman IA, Neto N, Papkovsky DB, Monaghan MG, Dmitriev RI: A deeper understanding of intestinal organoid metabolism revealed by combining fluorescence lifetime imaging microscopy (FLIM) and extracellular flux analyses. ***Redox Biology*** 2020, **30**:101420.
4. Peirsman A, Blondeel E, Ahmed T, Anckaert J, Audenaert D, Boterberg T, Buzas K, Carragher N, Castellani G, Castro F *et al*: MISpheroID: a knowledgebase and transparency tool for minimum information in spheroid identity. ***Nature Methods*** 2021. DOI: 10.1038/s41592-021-01291-4.
5. Debruyne AC, Okkelman IA, Heymans N, Pinheiro C, Hendrix A, Nobis M, Borisov SM, Dmitriev RI: Live Microscopy of Multicellular Spheroids with the Multimodal Near-Infrared Nanoparticles Reveals Differences in Oxygenation Gradients. ***ACS Nano*** 2024, **18**(19):12168-12186.

Development of intracellular phosphorescent O_2 probes for assessing cell hypoxia, cellular gradients and cell bioenergetics

As a postdoc, I contributed to advancing state-of-the-art of O_2 sensing based on the metalloporphyrin conjugates with cell-penetrating peptides and nanoparticles. These enabled studies of cell **bioenergetics** and **adaptation to hypoxia**, demonstration of **intracellular O_2 gradients in cell culture**, and indirectly catalyzed work and successful commercialization by colleagues inspiring a generation of researchers. This work and *state-of-the-art* are summarized in several high-impact review articles for the field (2012, 2013, 2015, 2018) and books (2012, 2018). This work also positively affected my career growth, helping to secure the follow-up funding from the Science Foundation Ireland (SFI, Starting Investigator grant 2014).

1. Dmitriev RI, Zhdanov AV, Ponomarev GV, Yashunski DV, Papkovsky DB: Intracellular oxygen-sensitive phosphorescent probes based on cell-penetrating peptides. ***Analytical Biochemistry*** 2010, **398:**24-33.
2. Dmitriev RI, Ropiak HM, Ponomarev GV, Yashunsky DV, Papkovsky DB: Cell-penetrating conjugates of coproporphyrins with oligoarginine peptides: rational design and application for sensing intracellular O_2**.** ***Bioconjugate Chemistry*** 2011, **22:**2507-2518.

3. Dmitriev RI, Zhdanov AV, Jasionek G, Papkovsky DB: Assessment of cellular oxygen gradients with a panel of phosphorescent oxygen-sensitive probes. ***Analytical Chemistry*** 2012, **84**(6):2930-2938.
4. Kondrashina AV, Dmitriev RI, Borisov SM, Klimant I, O'Brien I, Nolan YM, Zhdanov AV, Papkovsky DB: A phosphorescent nanoparticle-based probe for sensing and imaging of (intra)cellular oxygen in multiple detection modalities. ***Advanced Functional Materials*** 2012, **22**(23):4931-4939.

Developing multi-parametric FLIM of 3D tissue models and organoids

My research group in Cork (2014-2020) was among the first to demonstrate method for **monitoring the oxygenation of 3D tissue models** with the help of high-performance O_2-sensitive probes and PLIM method (Biomaterials, 2013; Biomaterials Sci 2014; CMLS 2014, Acta Biomater, 2015) and to combine it with the analyses of cell death, proliferation, cell temperature and markers of differentiation. Together with colleagues we also applied the FLIM and PLIM approaches to the **3D cancer spheroids** and **stem cell-derived intestinal organoid models**, demonstrating their utility for improved monitoring of their metabolism and viability (One, 2016, ACS Nano, 2015, Acta Biomater 2018, Adv Func Mater 2018). Currently (2023-present), we focus our efforts on visualizing polarity reversion in small intestinal organoids and its application to development of vaccines (2025), detection of micro- and nanoplastics (MNPs) and tissue engineering/ organ-on-a-chip applications:

1. Dmitriev RI, Kondrashina AV, Koren K, Klimant I, Zhdanov AV, Pakan JMP, McDermott KW, Papkovsky DB: Small molecule phosphorescent probes for O_2 imaging in 3D tissue models. ***Biomaterials Science*** 2014, **2**(6):853-866.
2. Dmitriev RI, Borisov SM, Düssmann H, Sun S, Müller BJ, Prehn J, Baklaushev VP, Klimant I, Papkovsky DB: Versatile Conjugated Polymer Nanoparticles for High-Resolution O_2 Imaging in Cells and 3D Tissue Models. ***ACS Nano*** 2015, **9**(5), 5275-5288.
3. Jenkins J, Borisov SM, Papkovsky DB, Dmitriev RI: Sulforhodamine nanothermometer for multi-parametric Fluorescence Lifetime Imaging Microscopy. ***Analytical Chemistry*** 2016, **88**(21):10566-10572.
4. Okkelman I. A., Dmitriev R. I., Foley T., Papkovsky, D. B.: Use of Fluorescence Lifetime Imaging Microscopy (FLIM) as a Timer of Cell Cycle S Phase. ***PLOS ONE*** 2016, **11**, e0167385.
5. O'Donnell N, Okkelman IA, Timashev P, Gromovykh TI, Papkovsky DB, Dmitriev RI: Cellulose-based scaffolds for fluorescence lifetime imaging-assisted tissue engineering. ***Acta Biomaterialia*** 2018, **80**:85-96.
6. *Sheehan A, Okkelman IA, Groslambert G, Bucher C, Dmitriev RI, Filatov MA: Optoelectronic Properties and Fluorescence Lifetime Imaging Application of Donor-Acceptor Dyads Derived From 2,6-DicarboxyBODIPY. ***Chemistry – A European Journal*** 2024, **n/a**(n/a):e202404188. **Cover image**: https://chemistry-europe.onlinelibrary.wiley.com/toc/15213765/2025/31/10
7. *Vanhoeijen R, Okkelman IA, Rogier N, Sedlačík T, Stöbener DD, Devriendt B, Dmitriev RI, Hoogenboom R: Poly(2-alkyl-2-oxazoline) Hydrogels as Synthetic Matrices for Multicellular Spheroid and Intestinal Organoid Cultures. ***Biomacromolecules*** 2025.

8. *Okkelman IA, Zhou H, Borisov SM, Debruyne AC, Lefebvre AEYT, Leomil Zoccoler M, Chen L, Devriendt B, Dmitriev RI: Visualizing the internalization and biological impact of nanoplastics in live intestinal organoids by Fluorescence Lifetime Imaging Microscopy (FLIM). ***Light: Science & Applications*** 2025, **14**(1):272.
9. *Vanhove L, Van Gansbeke T, Devriendt B, Van der Meeren R, Dmitriev RI, Okkelman IA: Lack of biochemical signalling in GelMA leads to polarity reversion in intestinal organoids independent from mechanoreciprocity**. *Journal of Tissue Engineering*** 2025, **16**:20417314251345000.
10. Chen L, Van Der Weken H, Zwaenepoel O, Okkelman IA, Aelvoet J, Van Denberghe E, Gettemans J, Dmitriev RI, De Geest BG, Cox E *et al*: Cell-specific mRNA delivery via nanobody-functionalized lipid nanoparticles. ***Journal of Controlled Release*** 2025, **388**:114365.

Contribution to the understanding of the mechanisms of cellular adaptation to the low oxygen environment (hypoxia), the stroke (oxygen and glucose deprivation).

This work was jointly performed with collaborators. Some of the main results are the demonstration of **augmented HIF signaling** under physiological O_2 levels in neuronal cells (Zhdanov et al, BBA, 2013), understanding of the **regulation of protein translation during stroke** (Andreev et al, Genome Biology, 2015) and discovery of the **ischemia-induced changes in histone acetylation** (Dmitriev, FEBS Lett 2015). The impact of these frontier research findings is expected to be in developing better therapies to prevent and improve survival of the tissue during stroke.

1. Zhdanov AV, Dmitriev RI, Golubeva AV, Gavrilova SA, Papkovsky DB: Chronic hypoxia leads to a glycolytic phenotype and suppressed HIF-2 signaling in PC12 cells. ***Biochimica et Biophysica Acta (BBA) - General Subjects*** 2013, **1830**(6):3553-3569.
2. Zhdanov AV, Dmitriev RI, Papkovsky DB: Bafilomycin A1 activates HIF-dependent signalling in human colon cancer cells via mitochondrial uncoupling. ***Bioscience Reports*** 2012, **32**:587-595.
3. Andreev D, O'Connor P, Zhdanov A, Dmitriev R, Shatsky I, Papkovsky D, Baranov P: Oxygen and glucose deprivation induces widespread alterations in mRNA translation within 20 minutes. ***Genome Biology*** 2015, **16**(1):90.
4. Dmitriev RI, Papkovsky DB: In vitro ischemia decreases histone H4K16 acetylation in neural cells. ***FEBS Letters*** 2015, **589**(1):138-144.
5. Barra J, Crosbourne I, Roberge CL, Bossardi-Ramos R, Warren JSA, Matteson K, Wang L, Jourd'heuil F, Borisov SM, Bresnahan E *et al*: DMT1-dependent endosome-mitochondria interactions regulate mitochondrial iron translocation and metastatic outgrowth. ***Oncogene*** 2024.

Molecular physiology and functional analysis of membrane ion-transporting proteins (mitochondrial transhydrogenase NNT and Secretory Pathway Calcium/ Manganese ATPase)

1. Arkblad EL, Tuck S, Pestov NB, Dmitriev RI, Kostina MB, Stenvall J, Tranberg M, Rydstrom J: A Caenorhabditis elegans mutant lacking functional nicotinamide nucleotide transhydrogenase displays increased sensitivity to oxidative stress. ***Free Radic Biol Med*** 2005, **38:**1518-1525.

2. Pestov NB, Dmitriev RI, Kostina MB, Korneenko TV, Shakhparonov MI, Modyanov NN: Structural evolution and tissue-specific expression of tetrapod-specific second isoform of secretory pathway Ca^{2+}-ATPase**.** ***Biochem Biophys Res Commun*** 2012, **417:**1298-1303.
3. Jenkins J, Papkovsky DB, Dmitriev RI: The Ca2+/Mn2+ -transporting SPCA2 pump is regulated by oxygen and cell density in colon cancer cells. ***Biochemical Journal*** 2016, **473**(16):2507-2518.

Please see my publications here:
Google Scholar https://scholar.google.com/citations?user=fAYmtSUAAAAJ&hl=en
University webpage: https://www.ugent.be/ge/hsr/en/research/tissue-engineering-and-biomaterials/overview.htm